

The PHILIPPINE JOURNAL OF

Veterinary Medicine

Volume 58

No. 1

January - June 2021

**Published by the College of Veterinary Medicine
University of the Philippines Los Baños**

ISSN 0031-7705

*The printing of this issue was made possible
through the support of the University of the
Philippines Veterinary Medicine Foundation, Inc.
and the Commission on Higher Education (CHED)
Journal Challenge Grant under its Journal
Incentive Program*

The Philippine Journal of Veterinary Medicine

Volume 58

No. 1

January-July 2021

The Philippine Journal of Veterinary Medicine is a peer-reviewed international journal of basic and applied research in veterinary medicine and science. It is published semi-annually, for the periods January-June and July-December each year, by the College of Veterinary Medicine, University of the Philippines Los Baños. All articles are subjected to double-blind review.

Authors of articles appearing in the journal are solely responsible for opinions expressed therein.

All rights reserved. No article of the journal may be reproduced in any form and by any means without a written permission from the publisher or the Editor-in-Chief.

EDITORIAL BOARD

Remil L. Galay, DVM, DVSc

Editor-in-Chief

Marianne Leila S. Flores, DVM, MHA

Associate Editor

Cherry P. Fernandez-Colorado, DVM, MS, PhD

Rio John T. Ducusin, DVM, MS, PhD

Michelle Grace V. Paraso, DVM, MSc, PhD

Gerry A. Camer, DVM, MS, PhD

Clarissa Yvonne J. Domingo, DVM, MPH, DrPH

Melbourne R. Talactac, DVM, MS, PhD

Technical Editors

Emilia A. Lastica-Ternura, DVM, MSc

Managing Editor

SUPPORT STAFF:

Junelle L. Paller

Fernando P. Micoso

Jocelyn E. Arcinas

Dino Mari F. Manasan

The annual subscription price is US\$100.00 (net) for foreign subscribers (inclusive of mailing cost) and Philippine PhP1,500.00 plus mailing cost for local subscribers. Prices for current single issue and back issues are available on request. Subscriptions are accepted on a prepaid basis only and are entered on a calendar year basis. Issues are sent by air delivery to foreign subscribers.

All communications should be addressed to:

The Editor-in-Chief

Philippine Journal of Veterinary Medicine

College of Veterinary Medicine

University of the Philippines Los Baños

Laguna, Philippines 4031

Telefax Nos. +63-49-536-2727, +63-49-536-2730

Email: pjvm1964@gmail.com, pjvm.uplb@up.edu.ph

This journal is abstracted/indexed by: SCOPUS, Biological Abstracts, Focus on: Veterinary Science & Medicine, Web of Science Zoological Records, CAB Abstracts, Index Veterinarius, Veterinary Bulletin, Parasitology Database, Helminthological Abstracts, Protozoological Abstracts, Review of Medical and Veterinary Entomology, EBSCO, ASEAN Citation Index, Prescopus Russia, *i-journals* (www.ijournals.my), *i-focus* (www.ifocus.my), *i-future* (www.ifocus.my), Philippine E-Journals (<https://ejournals.ph>) and UPLB Journals Online (<http://journals.uplb.edu.ph/index.php/PJVM>).

The Philippine Journal of Veterinary Medicine

Volume 58

No. 1

January – July 2021

CONTENTS

Original Articles

Diagnostic Imaging

- Ultrasound features of the liver, spleen, kidney and heart in dogs with canine parvoviral enteritis. 1
MM Mariño, JA Acorda, and AMGA Pajas

- Ultrasonographic features of the uterus and ovaries on Holstein-Sahiwal crossbred dairy heifers at different phases of the estrous cycle. 17
KYY Ponco, AMGA Pajas, and AA Rayos

Microbiology

- Phenotypic antimicrobial resistance patterns in *Escherichia coli* isolated from slaughtered healthy pigs and cattle in Nueva Viscaya, Philippines. 30
CD Bakakew, JV Tabuac, and HE Torio

- Isolation and uniplex polymerase chain reaction-based detection of *Salmonella* spp. in native chickens (*Gallus gallus domesticus* Linn.) from selected live bird markets in Batangas, Philippines. 40
JPF Galvez and DV Umali

- Virulence factor profile and antibiotic resistance of *Escherichia coli* O157 strains isolated from animal raw meat. 47
M Lofti, H Momtaz, and E Tajbakhsh

- Prevalence, phenotypic, and genotypic assessment of antibiotic resistance, virulence markers and molecular typing of *Staphylococcus epidermidis* strains isolated from bovine subclinical mastitic milk. 56
F Talebi, H Momtaz, and Z Bamzadeh

Parasitology

- First report of *Plagiorchis vespertilionis* (Müller, 1780), a known zoonotic fluke, with notes on two species of *Paralecithodendrium* (Platyhelminthes: Trematoda) from *Myotis* sp. and *Miniopterus* sp. (Mammalia: Chiroptera) in the Philippines. 70
SL Eduardo

Surgery

- Comparison of tiletamine-zolazepam-xylazine and ketamine-xylazine anesthesia in Philippine native goats undergoing rumenotomy
KRB Gicana, MJB Addatu, AMGA, Pajas, and JHA Abalos. 78

Zootechnics

- The relationship of body condition scores to milk production in dairy buffaloes. 84
TA Saludes, H Takeshita, AG Tandang, PM Baril, and JAN Bautista

Research Notes

Anatomy

- Histological characterization of the gut-associated lymphoid tissue in three-month old guinea fowls (*Numida meleagris*). 96
S Hamedi and M Shahmizad

Microbiology

- Serological and molecular detection of Newcastle disease in captive psittacines in a wildlife rescue center in Luzon, Philippines. 101
JA Baydo, EA Lastica-Ternura, and DV Umali

Case Report

- Mastitis in a Holstein x Sahiwal cow caused by streptomycin-resistant *Pasteurella multocida*. 108
RDO Manzanilla, and FMIR Pilapil-Amante

CASE REPORT**MASTITIS IN A HOLSTEIN X SAHIWAL COW CAUSED BY STREPTOMYCIN-RESISTANT *Pasteurella multocida***

Russel Denny O. Manzanilla, DVM
 Flor Marie Immanuelle R. Pilapil-Amante, DVM, MSc

*Department of Veterinary Clinical Sciences, College of Veterinary Medicine
 University of the Philippines Los Baños, 4031, Laguna, Philippines*

ABSTRACT

Mastitis, the most economically relevant disease in the dairy industry, can be caused by a wide array of bacterial species. Although scarcely reported, *Pasteurella multocida* is included as a contagious etiologic agent causing mastitis. Targeting the pathogens is the treatment approach when dealing with udder infection and with bacteria being the chief causative agents, antibiotics is prominently utilized for therapy. This dairy farm was previously treating its mastitis cases using a Penicillin-Streptomycin solution for intramuscular injection without veterinary supervision. The collection of milk samples from infected quarters was prompted by non-response of their mastitis cases to antibiotic treatment. The selection of quarters for sample collection was based on the severity of clinical signs present. Bacterial isolation was performed using the milk samples afterward, the isolated bacteria, *Staphylococcus aureus* and *P. multocida*, were subjected to antibiotic sensitivity tests. The two isolates shared a resistance against streptomycin, an antibiotic previously used in the farm in combination with benzylpenicillin. Thus, a different antimicrobial drug, oxytetracycline, was prescribed to which the pathogens responded positively.

Keywords: antimicrobial resistance, bovine mastitis, *Pasteurella multocida*, streptomycin

Philipp. J. Vet. Med., 58(1): 108-113, 2021

INTRODUCTION

With a decrease in milk production, an increase in veterinary expenses, and culling rates, bovine mastitis remains to be the most economically relevant disease in the dairy industry (Gomes, 2016). It is also considered as one of the most difficult veterinary diseases to control since over a hundred different microorganisms serve as the etiologic agent (Bradley, 2002). Mastitis may be identified by measuring the cell count in milk. An increase in the number of somatic cells in the milk is a result of an influx of polymorphonuclear (PMN) leucocytes from the mammary capillaries into the teat cistern as a response to bacterial proliferation (Blowey and Edmonson, 2010). Toxin production also contributes to mammary tissue damage and with increased permeability, hematologic elements are allowed to escape into the milk (Bekuma and Galmessa, 2018).

The pathogens responsible for mastitis include viruses, algae, and fungi but among these organisms, bacteria are reported to be the primary cause of bovine mastitis (Viguier, 2009). Contagious mastitis pathogens are transferred

from an infected cow to a non-infected one usually during milking (Blowey and Edmonson, 2010). *Staphylococcus aureus* was identified to be the most commonly involved mastitis organism as reported in most countries (Hossain, 2017). Environmental mastitis pathogens, on the other hand, originate from the surroundings with their infection rates vastly influenced by management practices (Blowey and Edmonson, 2010). As reported by Gomes *et al* (2016), *Streptococcus uberis* is the most frequently isolated environmental pathogen in bovine mammary infections. While there are new species becoming more commonly isolated, certain species remain to be uncommon causes of mastitis (Hamadani *et al.*, 2013).

Wilson and Ho (2013) describe *Pasteurella multocida* as a small, Gram-negative, non-flagellated coccobacillus. Isolates are preferably grown on 5% sheep's blood in dextrose-starch, casein-sucrose-yeast, chocolate, Mueller-Hinton, or brain-heart infusion agar and no growth can

***FOR CORRESPONDENCE:**

(e-mail: frpilapil@up.edu.ph)

be observed on MacConkey agar. *Pasteurella multocida* is the main causative agent in Hemorrhagic septicemia, in addition to its association with bovine pneumonia (Dabo *et al.*, 2007).

Among mastitis-causing pathogens, antimicrobial resistance is predominant in *Escherichia coli* and *Staphylococcus aureus*. It has been reported that generally, clinical mastitis cases exhibit resistance against penicillin, amoxicillin, oxytetracycline, and methicillin (Chandrasekaran *et al.*, 2014).

CASE PRESENTATION

In early January 2019, a dairy cattle farm comprised of a Holstein-Sahiwal milking herd with an average body condition score of 3/5 and located in Batangas, Philippines reported difficulties in the treatment of their mastitis cases. The animals that exhibited clinical signs were previously treated with Benzylpenicillin procaine-Benzylpenicillin benzathine-Dihydrostreptomycin sulphate combination (Pendistrep L.A., 120,000 IU/ 80,000 IU/ 200mg per ml solution; Phenix, Anupco Ltd., Essex, England) at a dose of 10 mL per 100 kg body weight through intramuscular injection once a day for 3 days without any supervision from a veterinarian. Upon consultation, the owner was advised to have milk samples from their mastitis-infected cows be collected and undergo bacterial isolation.

Physical examination and sample collection were performed on February 13, 2019, a month after the reported non-response to treatment. The caretakers emphasized that based from their California Mastitis Test (CMT), the number of their infected cows went up from five to thirteen. The said thirteen cows had at least one udder quarter with a grade of 3 from the CMT results based on standard scoring (Ruegg, 2005) which corresponds to >5,000,000 cells/ml and interpreted as severe mastitis infection. Sampling population was further reduced as only five cows, as shown in Figure 1A, exhibited mastitis clinical signs (hard and hot udder, coagulations in milk).

The yellowish milk samples (Fig. 1B) collected were taken to the Animal Disease Diagnostics Laboratory in the University of the Philippines Veterinary Teaching Hospital – Los Baños. Bacterial isolation was performed on the milk samples using Blood Agar Plate (BAP) and two bacterial species were isolated and cultured, *Staphylococcus aureus* and *Pasteurella multocida* (Table 1). Following bacterial growth after 48 hours, antibiotic sensitivity testing (against Norfloxacin, Florfenicol, Trimethoprim, Doxycycline, Tetracycline, Amoxicillin, Spectinomycin, and Streptomycin by Kirby-Bauer disk diffusion) was performed using Muller-Hinton agar and Blood Agar Plate, respectively. Results showed that the two bacterial species had developed intermediate resistance against

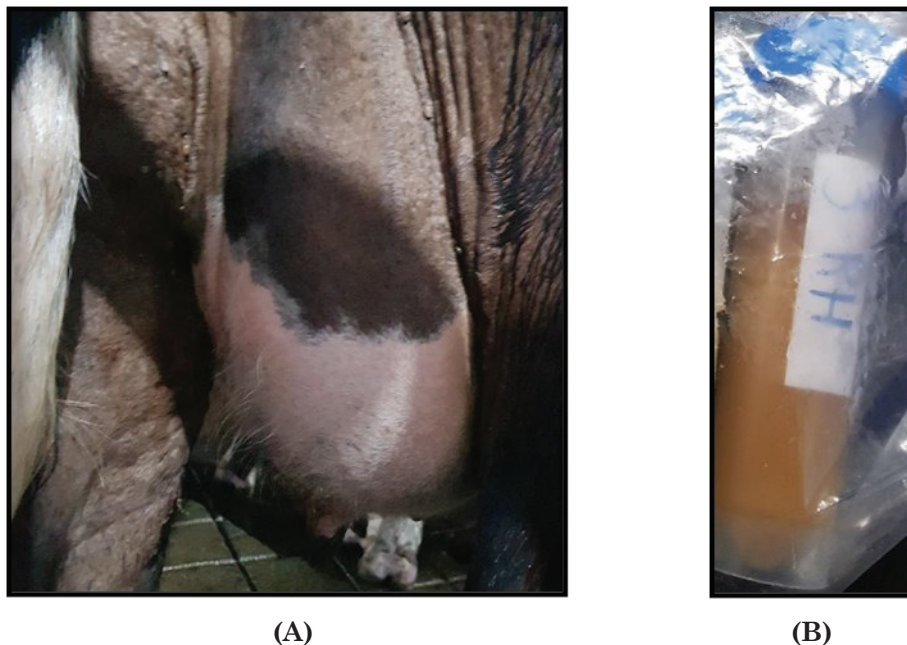


Fig 1. (A) Swollen appearance of cow # 3's mammary glands during milk sample collection; (B) Yellow milk sample collected from cow # 3.

Table 1. Bacterial isolation and Antibiotic Sensitivity Testing results (CLSI, 2013).

Sample ID	Results
Cow #1 LF	No growth after 48 hours of incubation
Cow #2 RH	No growth after 48 hours of incubation
Cow #3 RH	<p><i>Pasteurella multocida</i></p> <p>Sensitive: Norfloxacin, Florfenicol, Trimethoprim, Doxycycline, Tetracycline</p> <p>Intermediate: Amoxicillin, Spectinomycin</p> <p>Resistant: Streptomycin</p>
Cow #4 LF/RF	<p><i>Staphylococcus aureus</i></p> <p>Sensitive: Doxycycline, Florfenicol, Tetracycline</p> <p>Intermediate: Spectinomycin, Enrofloxacin, Norfloxacin</p> <p>Resistant: Streptomycin, Amoxicillin, Trimethoprim</p>
Cow #5 RF	No growth after 48 hours of incubation

Mammary Gland Quarters: LF Left front, RF Right front, LH Left hind, RH Right hind

Streptomycin. *Pasteurella multocida* had also developed intermediate resistance against Amoxicillin while *Staphylococcus aureus* had developed the same against Enrofloxacin and Norfloxacin and full resistance against Amoxicillin and Trimethoprim. Upon examining these results, the veterinarian prescribed a different class of antibiotic, oxytetracycline (Oxitetraciclina 200 L.A., 200mg per ml solution for intramuscular injection; Invesa, Livisto, Barcelona, Spain) at 1ml per 10 kg body weight once a day and to be repeated after three days as recommended by the manufacturer. Clusters of the milking machines were also examined. After identifying that the isolated pathogens are contagious in nature, samples using sterile swabs were acquired on February 15, 2019 from five randomly selected clusters. No bacteria were isolated from the swab samples, further suggesting that the isolated samples from the milk may have been transmitted by other means such as the milker's hands or the flies. The farm owner complied with the change in antibiotic treatment and had positive results such as recovery from inflammation and reduction of CMT score to negative (N) the following week leaving only Cow # 3 with clinical mastitis. Second dosing of Oxytetracycline was then advised which resulted to a better response to treatment and patient recovery.

DISCUSSION

Mastitis continues to plague the dairy industry as its most costly disease causing economic losses through different factors: treatment costs, untimely culling, and preventive expenditures. In addition to a declining milk production, milk quality is also negatively affected. Reduced milk components such as lactose, milk fat, and casein lessens product viability resulting to more economic losses (DeGraves *et al.*, 1993).

Bovine mastitis usually manifests in a clinical form, displaying symptoms both systemically and locally in the udder of the infected animal and milk produced, which was apparent in this farm. Subclinical mastitis also exists in the form of increased somatic cell count (SCC) subsequently diminishing milk produced (Franca *et al.*, 2017). Infection is primarily a result of bacterial proliferation in the mammary gland. Its broad diversity of microorganisms involved can be categorized as either 'contagious' or 'environmental' pathogens (Bradley, 2002). Contagious pathogens contribute to majority of mastitis cases generally causing subclinical form of udder infection. In contrast, infection by environmental pathogens normally manifest as clinical mastitis (Blowey and Edmonson, 2010). However, at the time of evaluation in this farm,

majority of the mastitis cases exhibited subclinical forms with more than half of the infected cows having quarters with a high SCC despite the absence of clinical signs.

One of the isolated organisms in this case is *P. multocida*, an opportunistic pathogen normally found in the oral, nasopharyngeal, and upper respiratory tract (Davies *et al.*, 2004). In a previous case report by Milanov *et al.* (2017), it was summarized that mastitis cases caused by *P. multocida* have a low incidence rate of not more than 1.25%; and data was retrieved only from a handful of studies. Immunosuppression, brought about by environmental stressors or nutritional deficiency, of the animal would allow *P. multocida* to proliferate and cause diseases in bovine such as Hemorrhagic Septicemia and Pneumonic Pasteurellosis (Davies *et al.* 2004). However, in cases of mastitis, the route of infection continues to be unclear, but several courses have been hypothesized. From the respiratory tract, organisms may be distributed via hematogenous or lymphogenic vessels into the udder. Suckling calves that have been infected by *P. multocida* may also serve as vectors as they may transfer the organism from their oropharynx to the udder while feeding (Milanov *et al.*, 2017).

Preventive measures against mastitis such as teat disinfection before and after milking and thorough cleaning of the milking system are practiced in this farm however, even with such control procedures, cows still develop mastitis eventually necessitating antibiotic treatment (Oliver and Murinda, 2012). Antibiotic therapy in mastitis cases is applied in two approaches: treatment of clinical infections during milking usually through intramammary administration and in severe cases, systemic antibiotics are used to supplement the treatment prior to drying off which is roughly six weeks before the next calving and the other technique, referred to as dry cow therapy, is the infusion of the udder with antibiotics (Krömker and Leimbach, 2017).

A 2008 study on antimicrobial susceptibility against *P. multocida* by Yoshimura *et al.* used minimum inhibitory concentrations (MIC) to compare the effectiveness of certain antibiotics against the said organism. The study included three antibiotics mentioned in this case and among them, the article reported that benzylpenicillin appeared to be the most active against *P. multocida*. Between the two other drugs, oxytetracycline performed better than dihydrostreptomycin with the latter having 19.4% of the isolates already resistant against the aminoglycoside. Even if penicillin was not included in the antibiotic sensitivity testing of

this case, another antibiotic, amoxicillin, belonging to the same drug family, was tested to which *P. multocida* had developed intermediate resistance. Although the study used a different method, their results agree with the pathogen resistance against streptomycin of this case.

Antimicrobial usage is considered as the principal source for pathogens' development of drug resistance, reinforced by administration of suboptimal concentrations and extra-label use of antibiotics (Sharma *et al.*, 2018). In addition to grower farms using antibiotics as growth promotants, non-adherence to the prescribed protocol, and excessive use of antibiotics to treat diseases (Pilapil-Amante *et al.*, 2020), the presented case may serve as a prime example of inappropriate use of antimicrobials as the dairy farm was previously treating its mastitis cases using an antibiotic in the absence of veterinary consultation thus, contributing to the budding problem about antimicrobial resistance. In practice today, antibiotic sensitivity testing is primarily done to affirm if the current therapy is appropriate or adjustments are necessary (Kromker and Leimbach, 2017). Further recommendations to the farm to improve the mastitis prevention, control, and management would include adopting a better cleaning and disposal program applicable to all farm areas to reduce and control the fly population (Levesque, 2004), practicing dipping of the milkers' hands in a disinfectant solution in between milking cows, and availability of written treatment plans, treatment, and disinfection records (Pilapil-Amante *et al.*, 2020).

ACKNOWLEDGEMENT

The authors would like to extend their gratitude to Dr. Ma. Adena J. Detera, who accompanied and guided the primary author during his visits to the dairy farm.

STATEMENT OF COMPETING INTEREST

The authors have no competing interests to declare.

AUTHOR'S CONTRIBUTION

The primary author contributed to the investigation, data curation and analysis, resources, visualization and original manuscript writing. The second author contributed to the conceptualization, investigation, methodology, data analysis, resources, project administration, supervision, visualization and manuscript review and editing.

REFERENCES

- Bekuma, A and Galmessa, U. 2018. Review on Hygienic Milk Products Practice and Occurrence of Mastitis in Cow's Milk. *Agricultural Research and Technology: Open Access Journal*. 18. 10.19080/ARTOAJ.2018.18.556053.
- Blowey R, and Edmondson P. 2010. Mastitis Control in Dairy Herds. 2nd Edition. CABI Publishing, CAB International, Oxfordshire, UK.
- Bradley A. 2002. Bovine Mastitis: An Evolving Disease. *The Veterinary Journal* 164 (2): 116-128.
- Chandrasekaran D, Venkatesan P, Tirumurugaan K, Nambi A, Thirunavukkarasu P, Kumanan K, Vairamuthu S and Ramesh S. 2014. Pattern of Antibiotic Resistant Mastitis in Dairy Cows. *Veterinary World* 7(6): 389-384.
- CLSI, 2013. Performance Standards for Antimicrobial Disk and Dilution Susceptibility Tests for Bacteria Isolated From Animals; Second Informational Supplement. CLSI document, VET01-S2. Clinical Laboratory Standards Institute, Wayne, PA, USA.
- Dabo S, Taylor, J and Confer, A. 2007. *Pasteurella multocida* and bovine respiratory disease. *Animal Health Research Reviews* 8 (02): 129–150.
- Davies, R, MacCorquodale R and Reilly S. 2004. Characterisation of bovine strains of *Pasteurella multocida* and comparison with isolates of avian, ovine and porcine origin. *Veterinary Microbiology* 99(2): 145–158.
- DeGraves, F and Fetrow, J. 1993. Economics of Mastitis and Mastitis [Control. *Veterinary Clinics of North America: Food Animal Practice*, 9(3), 421–434.
- França M, Del Valle T, Campana M, Veronese L, Nascimento G and Morais, J 2017. Mastitis causative agents and SCC relationship with milk yield and composition in dairy cows. *Archivos de Zootecnia* 66(253): 45-49.
- Gomes F, Saavedra M and Henriques M. 2016. Bovine mastitis disease/pathogenicity: evidence of the potential role of microbial biofilm. *Pathogens and Disease* 74.
- Hamadani H, Khan AA, Banday MT, Ashraf I, Handoo N, Bashir A and Hamadani A. 2013. Bovine Mastitis - A Disease of Serious Concern for Dairy Farmers. *International Journal of Livestock Research* 3(1): 42-55.
- Hossain M, Paul S, Hossain M, Islam M and Alam M. 2017. Bovine Mastitis and Its Therapeutic Strategy Doing Antibiotic Sensitivity Test. *Austin Journal of Veterinary Science & Animal Husbandry* 4(1): 1030.
- Krömker V and Leimbach S. 2017. Mastitis treatment-Reduction in antibiotic usage in dairy cows. *Reproduction in Domestic Animals* 52: 21-29.
- Levesque P. 2004. Less mastitis, better milk. Hoard's Dairyman. Institut de technologie agroalimentaire, Quebec, Canada.
- Milanov D, Aleksic N, Todorovic D and Bugarski D. 2017. *Pasteurella multocida* Mastitis in cow - Case Report. *Veterinarski Glasnik* (2): 117-122
- Oliver S and Murinda S. 2012. Antimicrobial Resistance of Mastitis Pathogens. *Veterinary Clinics of North America: Food Animal Practice* 28(2): 165-185.
- Pilapil-Amante FMIR, Baldrias LR, Rayos AA, Divina BP. 2020. Risk factor analysis on bovine mastitis in dairy herds of Batangas, Philippines. *Philippine Journal of Veterinary Medicine* 57(1): 41-53.
- Ruegg, PL. 2005. California mastitis test (CMT) fact sheet 1. Erişim Adresi:[<http://milkquality.wisc.edu/wp-content/uploads/2011/09/california-mastitis-test-fact-sheet.pdf>]. Erişim Tarihi, 19, 2013.
- Rochal B, Mendonça D and Ribeiro J. 2014. Trends in Antibacterial Resistance of Major Bovine Mastitis Pathogens in Portugal. *Revista Portuguesa de Ciências Veterinárias* 109: 79-88.
- Sharma C, Rokana N, Chandra M, Singh B, Gulhane R, Gill J, Ray P, Puniya A and Panwar H. 2018. Antimicrobial Resistance: Its Surveillance, Impact, and Alternative Management Strategies in Dairy Animals. *Frontiers in Veterinary Science* 4: 237.
- Viguier C, Arora S, Gilmartin, N, Welbeck K. and O'Kennedy R. 2009. Mastitis detection: current trends and future perspectives. *Trends in Biotechnology* 27(8): 486-493.
- Wilson BA and Ho M. 2013. *Pasteurella multocida*: from Zoonosis to Cellular

Microbiology. *Clinical Microbiology Reviews* 26(3): 631-655.

Yoshimura H, Ishimaru M, Endoh, Y, and Kojima A. 2008. Antimicrobial Susceptibility of *Pasteurella multocida* Isolated from Cattle and Pigs. *Journal of Veterinary Medicine, Series B*, 48(7): 555-560.